Evaluation of Antioxidant and Antidiabetic Activity of Mentha arvensis Linn.

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Mallikarjuna reddy kaipu, Research scholar, JNTUK, Kakinada, AP. Dr Manohar babu Sitty, Professor & Principal, SIMS College of Pharmacy, Mangaladas nagar, Guntur, AP.

Abstract:

The anti-diabetic activity of ethanol (70%) extract of *Mentha arvensis* was performed on *in vivo* model along with antioxidant activity and determination of total polyphenol content. Ethanol extract of *Mentha arvensis* contains 4.7% of total extractable polyphenols. The antioxidant activity studied on ethanol extract showed very promising result in superoxide radical, hydroxy radical and DPPH radical methods. The antioxidant activity is directly correlated to the anti-diabetic activity of drug. The in vivo anti-diabetic activity of ethanol extract of *Mentha arvensis* was performed on streptozotocin-induced diabetes mellitus showed significant inhibition of blood glucose level as compared to control and similar to that of standard glibenclamide. The overall data potentiates the traditional value of *Mentha arvensis* as an anti-diabetic drug.

Key words: Anti-diabetic, polyphenol, antioxidant, Mentha arvensi

Introduction:

Plants always play an important role in mankind life. Human beings use plants for their daily basic requirements like feeding, clothing, sheltering, hunting and nursing. As source of medicines, plants have formed the basis for sophisticated traditional systems and continue providing mankind with new remedies in treating different ailments. *M. arvensis* is a branched, strongly aromatic herb that stems up to 40 cm long with ascending terminal branches. The characteristics of Leaves are elliptic to oblong-ovate, 1.5 to 2 cm long, short-petioled, toothed margins, rounded or blunt tipped. Hairy purplish to light blue axillary flowers [1]. The plant is used to treat liver and spleen diseases, asthma and jaundice. The yield of oil is 5% by distillation of leaves which contain 40 to 50% menthol. The oil is used as antiseptic, carminative, refrigerant, stimulant and diuretic. Menthol, the main active constituent of *M. arvensis* is used in medicine for stomach disorders and in ointments for headache. The infusion of leaves is used for treatment of indigestion and rheumatic pains [2].

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Diabetes mellitus is a metabolic disorder characterized by hyperglycemia and changes in carbohydrate, fat and protein metabolism, associated with fixed or relative deficiencies in insulin secretion and or insulin action. it is currently an increasing global health concern. The incidence and prevalence of diabetes are growing, especially in developing and newly industrialized countries. Majority of all cases of diabetes in developed and developing countries are non-insulin-dependent diabetes mellitus, also known as type-2 diabetes (T2D), or adult-onset diabetes. These diagnoses are typically in adults more than 30 years of age [3], and are usually characterized by postprandial hyperglycemia, an abnormal rise in blood sugar following a meal [4]. Several factors integral to the T2D disease process are such as insulin resistance, hyperinsulinemia, impaired insulin secretion, reduced insulin-mediated glucose uptake and utilization [5]. Many efforts have been made to search for other effective and safe medicines from natural materials in order to control diabetes [6].

Indian traditional medicines have a long history to treat DM by the herbs and herbal extracts. *Mentha arvensi* is one of the traditional antidiabetic herbs. So far, the anti-diabetic activity of ethanol (70%) extract of *Mentha arvensis* (MAEE) was not performed on *in vivo* model. This study aimed to brief the antidiabetic properties of *Mentha arvensi* by correlating its antioxidant properties with *in vitro* and *in vivo* management of diabetes [7].

Materials & Methods:

Plant material and preparation of extract:

Leaves of *M. arvensis* plant were collected from Seshachalam hills and authenticated by Dr. K. Madhava Chetty, Assistant Professor in Department of Botany, Sri Venkateshwara University, Tirupati, Chittoor district, Andhra Pradesh. The crude plant material was dried under shade and powdered mechanically to coarse powder. The coarsely powdered plant material (500g) was subjected to extraction with 70% ethanol using soxhlet extraction. The extracts were concentrated to dry residue by distillation (temperature 60 °C without vacuum) and dried completely in desiccators and weighed. The yield of the ethanol extract was 4.6 g.

Phytochemical Screening: Standard screening tests [08] were employed in screening the extracts for different constituents. Conventional protocol for detecting the presence of alkaloids, tannins, flavonoids and steroids, etc. was utilized. The ethanol extract on TLC over silica gel has showed eight distinct spots, whereas hexane, ethyl acetate and hydroalcohol extracts of each has showed four to five spots. All the extracts were subjected to determination of total polyphenol content.

Animals:

Healthy adult Wistar Albino rats of 180-250 g of were selected for studying the anti-diabetic activity. The animals were obtained from Gentox laboratories, Hyderabad. The animals were housed according to CPCSEA guidelines (under standard temperature condition). They were given a pellet diet and water *ad libitum*. The ethical clearance was obtained from Institutional Animal Ethics Committee (IAEC) before the experiment.

Determination of total phenolic content in different extracts:

Preparation of standard: The total phenolic content in the plant extracts was determined by using Folin-Ciocalteu colourimetric method based on oxidation-reduction reaction [09]. Various concentrations of gallic acid solutions in methanol (10, 25, 50 and 75 μg/ml) were prepared. In a 20 ml test tube, 1 ml gallic acid of each concentration was added and to that 5 mL of Folin-Ciocalteu reagent (10%) and 4 ml of 7% Na₂CO₃ were added to get a total volume of 10 ml. The blue coloured mixture was shaken well and incubated for 30 minutes at 40 °C in a water bath and the phenols were determined by spectrophotometric method. The absorbance was measured at 765 nm against blank. All the experiments were carried out in triplicate and the mean value of absorbance was obtained. The average absorbance values obtained at different concentrations of gallic acid were used to plot the standard curve. The total phenolic content was expressed in terms of gallic acid equivalent (mg GA/g of extract), which is a common reference compound.

Preparation of sample: Various concentrations of the extracts (25, 50, 100 and 200 μ g/mL) were prepared. Following the procedure described for standard, absorbance for each concentration of extract was recorded. Total phenolics content of the extracts was expressed as mg gallic acid equivalents (GAE) per gram of sample in dry weight (mg/g). The total phenolic content in all samples was calculated using the formula: C = c V/m where, C = total phenolic content mg GAE/g dry extract, c = total phenolic acid obtained from calibration curve in mg/mL, V = total volume of extract in ml, total mass of extract in gram.

Statistical analysis: All the experiments were carried out in triplicates and data reported are mean \pm standard deviation. Calculation of linear correlation coefficient and correlation analysis were carried out using MS Office Excel 2007. The linear regression equation for a straight line is, Y = mx + c where, Y = absorbance of extract, m = slope of the calibration curve, x = concentration of extract, c = intercept. Using this regression equation, concentrations of extracts were calculated. From the calculated values of concentration of each extract, the total phenolics content was calculated.

Drugs: 1, 1- diphenyl-2-picrylhydrazyl (Sigma Chemical Company, St. Louis, USA), Riboflavin (Loba Chemie Pvt Ltd., Bombay), Deoxyribose (Sisco Research Laboratories Pvt Ltd., Mumbai), Nitroblue tetrozolium (Sisco Research Laboratories Pvt Ltd., Mumbai), All other chemicals and reagents used were of analytical grade, Ascorbic acid was utilized as reference antioxidant drug.

Determination of Superoxide Radical Scavenging Activity: Superoxide anion scavenging activity of ethanol extract of *M.arvensis* was done based on the method described by Liu, Ooi, and Hang [10] with slight modification. One millilitre of nitroblue tetrazolium (NBT) solution (156 mmol/L NBT in 100 mmol/L phosphate buffer, pH 7.4), 1ml NADH solution (468 mmol/L in 100 mmol/L phosphate buffer, pH 7.4) and 0.1 ml of sample solution of different concentrations of ET extracts in water were mixed. The reaction started by adding 100 ml of phenazine methosulphate (PMS) solution (60 mmol/L PMS in 100 mmol/L phosphate buffer, pH 7.4) to the mixture. The reaction mixture was incubated at 25°C for 5 min, and the absorbance at 560 nm was measured against blank samples. Decreased absorbance of the reaction mixture indicated increased superoxide anion scavenging activity. The percentage inhibition of superoxide anion generation was calculated using the following

Formula: Inhibitory ratio = $[(A_0-A_1)/A_0] \times 100$

Where A_0 was the absorbance of the control, and A_1 was the absorbance of the ET extracts and standards [11].

Determination of Hydroxyl Radical Scavenging Activity: Hydroxyl radical scavenging activity was measured by studying the competition between deoxyribose and the ethanol extract of *M.arvensis* for hydroxyl radicals generated from the $Fe^{2+}/EDTA/H_2O_2$ system (Fenton reaction). The hydroxyl radical attacks deoxyribose, which eventually results in the formation of thiobarbituric acid reacting substances (TBARS) [12]. Fenton reaction mixture consisting of 200 μl of 10 mM ferrous sulphate (FeSO₄. 7H₂O), 200 μl of 10 mM EDTA and 200 μl of 10 mM 2-deoxyribose and was mixed with 1.2 ml of 0.1 M phosphate buffer (pH 7.4) and 200 μl of plant extract. Thereafter, 200 μl of 10 mM H₂O₂ was added before the incubation at 37 °C for 4 h. Then, 1 ml of this Fenton reaction mixture was treated with 0.2 ml of 8.1% sodium dodecyl sulphate, 1.5 ml of 0.8% thiobarbituric acid and 1.5 ml of 20 % acetic acid. The total volume was then made to 5 ml by adding distilled water and kept in an oil bath at 100 °C for 1 hour. After the mixture had been cooled, 5 ml of 15:1 v/v butanol-pyridine mixture was added. Following vigorous shaking, the

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tubes were centrifuged at 4000 rpm for 10 min and the absorbance of the organic layer containing the thiobarbituric acid reactive substances was measured at 532 nm. A control was prepared using 0.1 ml of vehicle in the place of plant extract/ascorbic acid. The percentage inhibition of hydroxyl radicals by the extract/compound was determined by comparing the absorbance values of the control and the experimental tubes as calculated for hydroxyl radical assay.

The percentage inhibitions of superoxide production by the extracts were calculated using the formula: Inhibitory ratio = $[(A_0-A_1)/A_0] \times 100$

Where, A₀ is the absorbance of control; A₁ is the absorbance with addition of plant extract/ ascorbic acid.

Determination of 1, 1- Diphenyl-2-Picrylhydrazyl (DPPH) Radical Scavenging Activity: The free radial scavenging activity of ethanol extract of *M.arvensis* was measured in terms of hydrogen donating ability using DPPH radical as described by the method [13] with a slight modification. Briefly, 40 μ l of sample extracts of different concentrations (0.05–2 mg/ml) were mixed with 200 μ l of 50 μ M DPPH solution in ethanol. The mixture was immediately shaken and incubated for 15 min in the dark at room temperature. The decrease in absorbance was measured at 517 nm with a microplate reader (Tecan Sunrise, Austria). Ascorbic acid (5–80 μ g/ml) was used as a standard and the control was ethanol. The percentage of inhibition activity of the extracts was calculated according to the following equation:

Percent (%) inhibition of DPPH• activity = $[(A_0-A_1)/A_0] \times 100$

Where, A₀ is the absorbance of control; A₁ is the absorbance with addition of plant extract/ ascorbic acid.

The concentration of extracts required to scavenge 50% of DPPH radical was estimated from the graph plotted against the percentage inhibition and compared with the standard. All the tests were performed in triplicate, and the results were expressed as $\mu g/ml$.

Calculation of 50% inhibition concentration: The optical density obtained with each concentration of the extract/ascorbic acid was plotted taking concentration on X-axis and percentage inhibition on Y-axis. The graphs were extrapolated to find the 50% inhibition concentrations of extracts/ascorbic acid.

Anti-diabetic activity:

Effect of ethanol extract of *M. arvensis* on oral glucose tolerance test:

Wistar Albino rats (healthy) were selected and randomly divided into seven groups (n=6). Group 'I' serves as normal control receiving distilled water, Group 'II', 'III', 'IV', 'V' serves as test groups receiving MERB (150 mg/kg b.w., and 300 mg/kg b. w.) and EARB (150 mg/kg b.w.and 300 mg/kg b. w.), Blood was withdrawn from the retro orbital plexus at 60, 90, 120 and 150 minutes after 1 h of drug administration.13,14 The glucose level in blood was measured by using GOD-POD method.

Effect of ethanol extract of *M. arvensis* instreptozotocin induced diabetic rats:

The whole study was divided into five groups, each group contain six animals. Streptozotocin (60 mg/kg) was injected to rats in each group of animals following 18 h fast. After 48 h the blood glucose level of rats were determined by glucose oxidase method. Only those animals which showed hyperglycaemia (blood glucose levels more than 200 mg/dl), were considered diabetic and taken for further experimentation. The group I received vehicle only (Tween-80, 1%) and served as control. Animals of group II, group III and group IV received MAEE 100, 200 and 400 mg/kg body wt therapeutic doses, respectively and group V received glibenclamide 1.0 mg/kg. The study was carried out after repeated (once a day) administration of the extract for 21 consecutive days. Basal glycaemia was determined in overnight fasted animals. The vehicle (1% tween 80), MAEE (100, 200 and 400 mg/kg) and

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glibenclamide (1.0 mg/ kg) were orally administered to the animals of group I-V for 21 consecutive days. Blood samples collected on the 7th, 14th and 21st days were analyzed for the determination of blood glucose level [14].

Statistical Analysis: Results were analyzed using one way ANOVA method and expressed as Mean \pm SEM. The statistical significance considered was P < 0.05 (confidence limit: 95%).

RESULTS AND DISCUSSION:

In the present study we have studied phytochemical screening, total phenolic content and *in vitro* anti-oxidant and anti-diabetic activity of ethanol extract of *M. arvensis* leaves. Phytochemical screening revealed the presence of tannins, flavonoids and triterpenoids. The total phenolic content of ethano extract is 379.36 ± 6.41 mg/g GAE. Total phenolic content of the extracts was calculated from the regression equation of calibration curve (y = 0.014x; $R^2 = 0.999$) and expressed as mg gallic acid equivalents (GE) per gram of sample in dry weight (mg/g).

Superoxide anion plays an important role in the formation of more reactive species such as hydrogen peroxide, hydroxyl radical and singlet oxygen, which induce oxidative damage in lipids, proteins and DNA [15]. Therefore, studying the scavenging activity of plant extracts/compounds on superoxide radical is one of the most important ways of clarifying the mechanism of antioxidant activity. In the present study, ethanol extract of *M.arvensis* was found to possess concentration dependent scavenging activity on superoxide generated by photoreduction of riboflavin and the results were graphically shown in Figure 1.

The mean 50% inhibition concentration (IC $_{50}$) values for superoxide radical of ethanol extract of *M. arvensis* was found to be 147.25 μg . The mean IC $_{50}$ value of ascorbic acid was found to be 52.5 μg . The results were given in Table 1 and Figure 4.

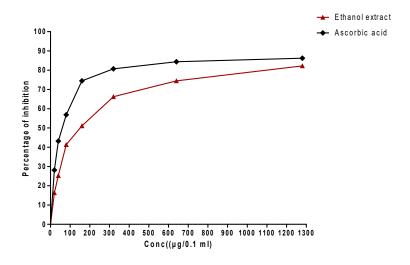


Fig 1: Concentration dependent percentage inhibition of Superoxide radical by ethanol extract of M.arvensis.

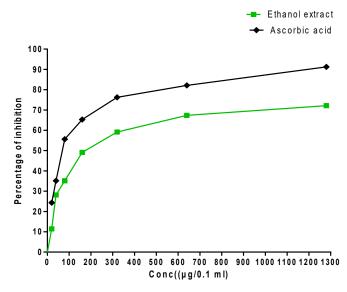


Fig 2: Concentration dependent percentage inhibition of Hydroxyl radical by ethanol extract of M.arvensis

The ethanol extract of M.arvensis was found to possess concentration dependent scavenging activity on hydroxyl radicals and the results were graphically shown in Fig 2. The mean IC₅₀ values for hydroxyl radical of ethanol extract of M.arvensis was found 181.22 μg . The mean IC₅₀ value of ascorbic acid was found to be 66.8 μg . The results were shown in Table 1 and Figure 4.

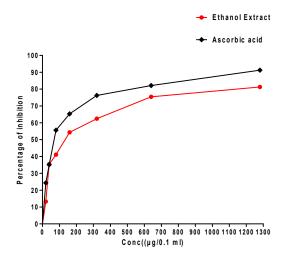


Fig 3: Concentration dependent percentage inhibition of DPPH radical by different extracts of M.arvensis.

The ethanol extract of M.arvensis was found to possess concentration dependent scavenging activity on DPPH radicals and the results were graphically shown in Fig 3. The mean IC₅₀ values for DPPH radical of ethanol extract of M.arvensis was found to be 116.42 μg . The mean IC₅₀ value of ascorbic acid was found to be 19.5 μg . The results were given in Table 1 and Figure 4.

Antioxidants plays a vital role in the prevention of human diseases. Herbal extracts with antioxidants activity may work as free radical scavengers, reducing agents and quenchers of single oxygen formation or reactive oxygen species, thereby protecting the health from degenerative disease such as cancer. The reactive oxygen species are damaging byproducts generated during normal cellular metabolism or from toxic injury. They lead to cause oxidative stress that contributes to the development of number of human diseases by damaging lipids, proteins and

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DNA. Phenolic content have shown a good correlation with antioxidant activity, this may be due to structural differences. Phenolic compounds, such as tannins, flavonoids and phenolic acids possess anti-inflammatory, anti-carcinogenic, anti-atherosclerotic and other properties that may be related to their antioxidant activities [16].

The ethanol extract of *M. arvensis* produced concentration dependent percentage inhibition on superoxide, hydroxyl radicals and DPPH radicals and produced maximum activity at a concentration of 160 μ g and there after the percentage inhibitions were raised gradually to its maximum level with higher concentrations.

Table 1: 50% Inhibition concentrations	(IC_{50}) of ethanol ex	xtract of <i>M.arvensis</i> against	Superoxide, Hydroxy	l and DPPH radicals.
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Extracts/Compound		50% Inhibition Conc (IC:	50)
Extracts/Compound	Superoxide radical	Hydroxyl radical	DPPH radical
M.arvensis Ethanol extract	146.25±1.3	181.22±3.1	116.42±3.2
Ascorbic acid	52.5±2.1	66.8±2.3	19.5±1.5

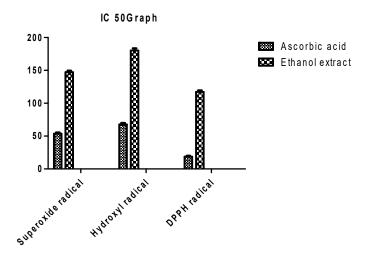


Figure 4: 50% Inhibition concentrations (IC50) of ethanol extract of M. arvensis against Superoxide, Hydroxyl and DPPH radicals.

In our study, induction of type 2 diabetes showed significant increase in blood glucose level and reduced body weight and insulin level compared to control rats which prove the induction of diabetes, and it may be due to partial necrosis of pancreatic β -cells [17]. Also, body weight of diabetic rats was reduced, and it may be due to decrease in amount of insulin. Physiologically, insulin regulates protein synthesis and proteolysis in .

In vivo study of MAEE was done on streptozotocin induced diabetic rats. The effect of the treatment with extract and glibenclamide on blood glucose level in diabetic rats after post-treatment days was shown in Table 3. The study was conducted by collecting blood on 7th, 14th and 21st days after induction of diabetes and analyzed for the determination of blood glucose level. All the values were found significant in the treatment doses (200 and 400 mg/kg body wt) as compared to control and standard glibenclamide (Table 3). These findings suggested that ethanol extract of *M.arvensis* has anti-diabetic activity, which validates its traditional uses in India.

Table 2: Effect of ethanol extract of M.arvensis on oral glucose tolerance in rats

Time in hours						
Treatment (mg/kg)	Fasting	1	2	3	4	5
Control	98.28±3.79	98.07±4.32	97.62±4.78	97.59±4.08	97.74±4.40	97.02±3.86
MAEE 100	114.77±3.43	109.32±6.62	103.36±5.21	99.62±4.64	97.49±5.15	99.39±4.32
MAEE 200	94.39±1.88	89.41±2.69	86.41±2.84	79.83±2.81	74.30±2.42	79.83±2.08
MAEE 400	93.3±1.54	84.9±1.74	79.49±2.55	73.36±3.33	68.38±2.28	71.94±1.61

^{*} Significant at p < 0.05, ** Significant at p < 0.01, *** Significant at p < 0.001, NS Not significant compared to control

Table 3: Effect of repeated oral administration of Ethanol extract of *M. arvensis* (MAEE) and glibenclamide on blood glucose level in streptozotocin diabetic rats.

Treatment	Blood glucose profile (mg/dl)			
	0	7	14	21
Control (vehicle)	342.20±15.01	367.00±24.31	395.00±24.31	419.00±24.44
MAEE 100 mg/kg	337.05±11.53	321.05±10.74	307.05±11.53	291.05±11.53
MAEE 200 mg/kg	321.00±21.46	272.80±13.60*	244.80±16.60*	209.40±13.55*
MAEE 400 mg/kg	319.00±16.06	245.80±16.60*	219.80±16.60*	181.40±13.55*
Glibenclamide 1 mg/kg	291.00±9.44	221.00±17.84*	184.70±15.84*	164.00±11.79*

The values represent in±SEM, n=6, *<0.05.

All the extracts of *M. arvensis* roots in this research exhibited different extent of antioxidant activity in different assays. It is evident from the present study that the ethanolic extract of *M. arvensis* leaves could be used as good source of natural potent antioxidants in controlling *Diabetes millitus*.

CONCLUSION:

The phytochemical screening tests indicated the presence of tannins, flavonoids and triterpenoids in the ethanol extract of *M.arvensis*. Several of such compounds are known to possess potent antioxidant and anti-diabetic activities [18]. Hence, the observed antioxidant and anti-diabetic activities may be due to the presence of any of these constituents. Further purification of these bioactive constituents may result in the development of potent antioxidant agent with low toxicity and better anti-diabetic index.

CONFLICT OF INTEREST:

The authors declare that there is no conflict of interests regarding the publication of this paper.

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References:

- 1. Ellis BE, Towers GH (1970). Biogenesis of rosmarinic acid in Mentha. Biochem. J., 118: 291-297.
- Rathish N, Sumitra VC (2007). Antibacterial Activities of Some Medicinal Plants of the Western Region of India, Turk. J. Biol., 31:231-236.

ISSN NO: 2236-61242

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- 3. Sidik SM, Ahmad R. Dietary management of a patient with diabetes mellitus: a case report. Mal J Nutr. 2003; 9(2): 137–144.
- 4. Apostolidis E, Kwon YI, Shetty K. Inhibitory potential of herb, fruit, and fungal-enriched cheese against key enzymes linked to type 2 diabetes and hypertension. Innov Food Sci Emerg Technol. 2007; 8(1): 46–54.
- 5. Tiwari AK, Madhusudana Rao J. Diabetes mellitus and multiple therapeutic approaches of phytochemicals: Present status and future prospects. Curr Sci. 2002; 83(1): 30–38.
- Matsui T, Ueda T, Oki T, Sugita K, Terahara N, Matsumoto K. α-Glucosidase inhibitory action of natural acylated anthocyanins 1. Survey of natural pigment with potent inhibitory activity. J Agric Food Chem. 2001; 49(4): 1948–1951.
- 7. Grover JK, Yadav S, Vats V. Medicinal plants of India with antidiabetic potential. J Ethnopharmacol 2002;81:81-100.
- 8. Trease GE and Evans WC. A text book of Pharmacognosy. 13th edn. Bacilluere Tinal Ltd, London;1989.
- 9. Waterhouse A. Determination of total phenolics. In: Wrolstad RE, editor. Current protocols in food analytical chemistry. New York: John Wiley and Sons; 2002. pp. I.1.1.1–II.1.8.
- 10. Liu F, Ooi VEC, & Chang ST. Free radical scavenging activity of mushroom polysaccharide extracts. Life Science. 1997;60:763–771.
- 11. Ye XY, Wang HX, Liu F, & Ng TB. Ribonuclease, cell-free translation-inhibitory and superoxide radical scavenging activities of the iron-binding protein lactoferrin from bovine mill. Int. J.Biochem.Cell Biol. 2000;32:235–241.
- 12. Elizabeth kunchandy and Rao MN. "An Oxygen radical scavenging activity of curcumin". Int. J. Pharm. 1990; 58:237-40.
- 13. LA Marghitas, OG Stanciu, DS Dezmirean. In vitro antioxidant capacity of honeybee-collected pollen of selected floral origin harvested from Romania. Food Chem. 2009;115(3):878–883.
- 14. Bergymeyer HU, Benot E. Colorimetric methods with glucose oxidase and peroxidase. In: Kunst A, Draeger B, Ziegenhorn J, Bergymeyer H, editor. Methods of enzymatic Analysis. 3rd ed. New York: Academic Press; 1963, p. 179-85.
- 15. Pietta PG. Flavonoids as antioxidants. J Nat Prod. 2000;63(7)1035-42.
- 16. Ravi Parimi*, K E Pravallika. Studies on Phytochemical Screening, Total Phenolic Content and In vitro Antioxidant activity of Different Extracts of *M. arvensis* Roots. *Research J. Pharm.* and Tech. 2017; 10(2): 551-556.
- 17. Dineshkumar B, Mitra A and Manjunatha M. A comparative study of alpha amylase inhibitory activities of common anti-diabetic plants at Kharagpur block. *International journal of Green Pharm* 2010; 4:115-21.
- 18. Lee J, Koo N, Min DB. Reactive oxygen species, aging and antioxidative neutraceuticals. Compr. Rev. Food Sci. Food Saf. 2004;3:21-33.